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1 **The crucial role of genome-wide genetic variation in conservation**

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32
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43 **Abstract**

44 The unprecedented rate of extinction calls for efficient use of genetics to help conserve
45 biodiversity. Several recent genomic and simulation-based studies have argued that the field of
46 conservation biology has placed too much focus on conserving genome-wide genetic variation,
47 and that the field should instead focus on managing the subset of functional genetic variation that
48 is thought to affect fitness. Here, we critically evaluate the feasibility and likely benefits of this
49 approach in conservation. We find that population genetics theory and empirical results show that
50 conserving genome-wide genetic variation is generally the best approach to prevent inbreeding
51 depression and loss of adaptive potential from driving populations towards extinction. Focusing
52 conservation efforts on presumably functional genetic variation will only be feasible occasionally,
53 often misleading, and counterproductive when prioritized over genome-wide genetic variation.
54 Given the increasing rate of habitat loss and other environmental changes, failure to recognize the
55 detrimental effects of lost genome-wide genetic variation on long-term population viability will
56 only worsen the biodiversity crisis.

57

58 **Introduction**

59 Decades of theoretical (1) and empirical (2, 3) research suggest that conserving genome-wide
60 genetic variation improves population viability. Maintaining genetic variation, *adaptive potential*
61 (see Glossary), and avoiding *inbreeding depression* are central motivations for maintaining large,
62 connected natural populations. Principles of genetics and evolution have therefore played a large
63 role in conservation biology since its inception (4, 5). The genomics revolution has inspired
64 biologists to leverage genome analysis to advance conservation beyond what was possible with
65 traditional genetics. Numerous studies have sequenced genomes of non-model organisms of
66 conservation concern to understand population history, *inbreeding* depression, and the genetic
67 basis of adaptation. A particularly exciting area of research has been to determine when and how
68 functional genetic information can advance conservation.

69 Several recent studies suggest that too much emphasis has been placed on genome-wide
70 genetic variation in conservation biology. For example, persistence of small populations for long
71 periods of time despite low genetic variation, and the collapse of the Isle Royale wolf population
72 after the infusion of genetic variation via immigration, have been interpreted as a challenge to the
73 idea that genetic variation generally increases population viability (6-12). Additionally, a weak
74 relationship between conservation status and genetic variation has been used to argue that genome-

75 wide (presumably neutral) genetic variation is of little importance to conservation (11). Several
76 authors have thus advocated for an approach that focuses on functional genetic variation that is
77 thought to directly affect fitness (including minimizing deleterious genetic variation) in place of
78 the traditional emphasis on conserving genome-wide genetic variation (6-8, 11).

79 Here, we evaluate the theoretical and empirical basis of this challenge to the importance of
80 genome-wide genetic variation and show that its premise is inconsistent with population genetic
81 theory and empirical findings. While it is clear that functional genetic information can advance
82 conservation, deemphasizing the maintenance of genome-wide genetic variation would increase
83 the extinction risk of threatened populations.

84

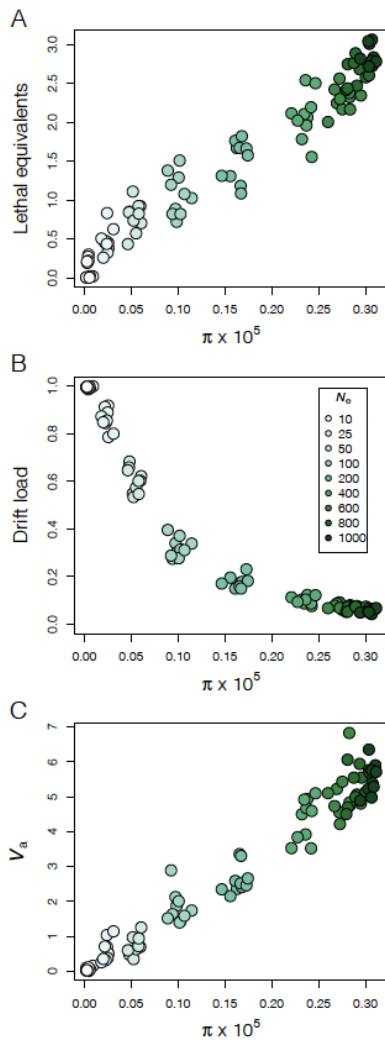
85 **1. Is genetic variation predictive of inbreeding and inbreeding depression?**

86 Inbreeding depression is thought to be driven mainly by homozygous and *identical-by-descent*
87 deleterious, partially recessive alleles (13), with lethal and small effect deleterious alleles
88 contributing substantially (14). The constant input of new deleterious mutations (15-19) makes
89 inbreeding depression a ubiquitous phenomenon that can push populations toward extinction (2,
90 20-23). One of the foundational predictions of theoretical population genetics is that the rate of
91 loss of heterozygosity (H) per generation ($\Delta\bar{H}=1/2N_e$) is identical to the rate of increase in mean
92 individual inbreeding (F), which is $\Delta\bar{F}=1/2N_e$ (24). \bar{H} is therefore expected to be entirely
93 predictive of \bar{F} (24-29).

94 A more difficult, but crucial question is whether genome-wide genetic variation (π) is
95 predictive of inbreeding depression. Deleterious alleles are lost in small populations due to
96 selection and genetic drift (30, 31), but they are also more often expressed in homozygotes in
97 smaller populations due to inbreeding. Selective *purging* of large effect deleterious alleles
98 following inbreeding combined with genetic drift may therefore result in low *inbreeding load* and
99 little inbreeding depression in the most highly inbred populations with the lowest π . However, the
100 presence of purging does not imply that high fitness is maintained in small populations with low π .

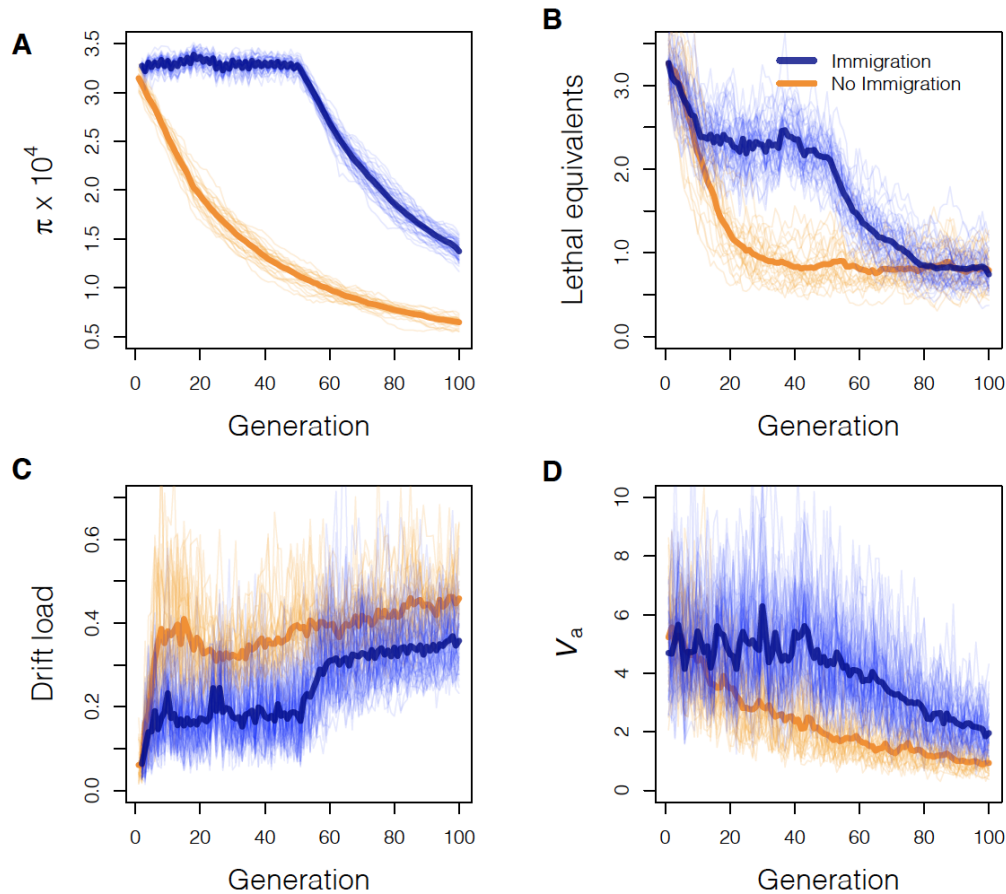
101 Population genetics theory predicts that larger populations will have higher neutral (24)
102 and deleterious genetic variation (32, 33). This is illustrated in Figure 1, where simulated large
103 populations have higher π (24) and higher inbreeding load (32-34) arising from segregating
104 partially recessive deleterious alleles. These simulations assume empirically supported models of
105 fitness and dominance (h) effects (*Supplementary Information [SI]*). Smaller populations have

106 lower π due to genetic drift, and fewer *lethal equivalents* due to genetic drift and purging.
107 However, despite having fewer lethal equivalents, chronically smaller populations have lower
108 mean fitness due to partially recessive deleterious alleles being expressed following inbreeding,
109 and some reaching high frequency or fixation (i.e., high *drift load*). Therefore, a negative
110 relationship is expected between π and drift load for populations at mutation-drift-selection
111 equilibrium.
112



113
114 **Figure 1.** Relationship of nucleotide diversity (π) with the inbreeding load (lethal equivalents)
115 (A), drift load (B), and additive genetic variance in a quantitative trait (V_a) (C). The data are from
116 the 1,000th generation of 10 simulated populations with 9 different constant effective population
117 sizes (N_e).

118 Equilibrium levels of π and drift load are not expected in populations with fluctuating
119 population size or immigration rate. A common scenario with high conservation relevance is
120 isolated populations that have experienced recent bottlenecks. The simulated data in Figure 2
121 shows that genome-wide π declines over time following a bottleneck, as expected from classical
122 theory (24) (Figure 2A). This pattern is paralleled by lethal equivalents (Figure 2B) owing to the
123 loss of deleterious alleles via genetic drift and purging of deleterious alleles expressed in
124 homozygotes due to inbreeding (30, 31). However, the deleterious alleles remaining after a
125 bottleneck often go to high frequency or fixation. This results in individuals being homozygous for
126 increasingly more deleterious alleles (higher drift load, Figure 2C) as π declines inexorably during
127 a sustained bottleneck, the same pattern expected for small populations at equilibrium (Figure 1).
128 It is notable, though, that π , inbreeding load, and drift load can change at substantially different
129 rates following a bottleneck. For example, drift load can become quite high before π declines
130 substantially following a bottleneck (Figure 2A, 2C). However, small populations that already
131 have low π are also expected to have low mean fitness due to ever-increasing drift load, which
132 demonstrates that π is a good indicator of drift load and mean fitness. Occasional immigration can
133 be sufficient to maintain high π and low drift load in small populations (Figure 2). This is one
134 reason why maintaining connectivity is a priority in conservation biology, and why *genetic rescue*
135 is an effective tool for managing small, isolated populations (30, 35, 36).



136
 137 **Figure 2.** Genetic effects of bottlenecks with and without immigration. Nucleotide diversity (π)
 138 (A), number of lethal equivalents (B), drift load (C), and the additive genetic variance in a
 139 quantitative trait (V_a) (D) are shown for 100 generations after a simulated bottleneck in isolated
 140 populations (orange) and with 5 immigrants every 2 generations up to generation 50 (blue).
 141 Population size was held constant at $N_e=1,000$ for 1,000 generations before the bottleneck and then
 142 at $N_e=25$ starting at generation 0. The thin lines show the results from 25 replicates. The thick lines
 143 represent the mean across 25 replicates. Immigrants during the first 50 generations are from a
 144 population with $N_e=500$ that split from the receiving population the generation of the bottleneck.
 145 Details of the simulation model and parameters are provided in the SI.

146
 147 Empirical data show that purging does not eliminate the extinction threat posed by
 148 inbreeding. Pedigree-based studies have yielded mixed results with regard to purging, with
 149 typically only a small portion of inbreeding depression being removed after sustained inbreeding
 150 in small populations (37-39). Analyses of 60 genomes from seven ibex species found that species
 151 which went through the most severe bottlenecks had more deleterious alleles (40). Alpine ibex,

152 which were once reduced to 100 individuals, had fewer highly deleterious alleles but more mildly
153 deleterious alleles compared to Iberian ibex (bottleneck size 1,000 individuals). Empirical genetic
154 data suggest small populations have higher drift load (40-42) which has resulted in lower
155 population growth in populations with lower genetic variation (2, 3). In agreement with
156 theoretical expectations outlined above, these data suggest that purging is insufficient to maintain
157 high fitness in the face of strong genetic drift and inbreeding. Thus, the presence of genomic
158 signatures of purging should not be taken as evidence for the absence of inbreeding depression, or
159 for demographic stability of small populations.

160 The relationship between π and fitness is obviously complicated, particularly immediately
161 after a bottleneck (Figure 2). Populations with the lowest π and highest inbreeding will also have
162 the lowest inbreeding load on average due to reduced deleterious genetic variation via genetic drift
163 and purging. However, these same genetically depauperate populations will typically have lower
164 fitness than larger, genetically diverse populations on average due to ever-increasing drift load
165 (Figures 1 & 2). The bottom line is that reduced fitness is generally expected in small, isolated,
166 genetically depauperate populations due to inbreeding depression and the accumulation of drift
167 load, and that maintaining genetic variation and population connectivity will increase long term
168 viability.

169

170 **2. Is genome-wide genetic variation predictive of adaptive potential?**

171 The ability of populations to adapt to changing environmental conditions (*adaptive potential*) is
172 fundamental for persisting through environmental change (43, 44). A core insight from theoretical
173 genetics is that adaptation requires additive genetic variance (V_a) for the selected trait(s) (45). A
174 lack of V_a can limit a population's response to selection and eventually lead to extinction (43, 44,
175 46). As with other types of genetic variation, V_a is affected by mutation at loci affecting the trait,
176 selection, migration, and genetic drift (47). We therefore expect from first principles that larger
177 populations will have higher π and higher V_a than small populations *on average* (Figure 1), and
178 thus that π should be correlated with V_a . Despite strong theoretical support, determining the
179 strength and importance of this relationship in real populations, especially those of conservation
180 concern, has generated longstanding controversy (48).

181 Basic population genetic theory shows that population size and connectivity play major
182 roles in determining V_a , and thus adaptive potential. Isolated populations below a certain size

183 should lose V_a due to genetic drift more rapidly than it is replenished via mutation (47).
184 Additionally, recently bottlenecked populations that have lost π will eventually also lose V_a and
185 evolutionary potential in the absence of immigration (Figure 2). However, while the eventual
186 reduction in V_a in small populations is inevitable, the initial effects of a bottleneck on V_a can be
187 complex. Recently bottlenecked populations may show decreases, stability, or even short-term
188 increases in V_a due to the conversion of dominant or epistatic variance into V_a as allele frequencies
189 change due to genetic drift (49-51). This potential conversion of nonadditive to additive variation
190 in bottlenecked populations is highly stochastic across traits and populations, and is one of the
191 processes that can cloud the relationship between molecular and quantitative trait variation (52).
192 Nonetheless, the two important takeaways are: 1) although bottlenecks can complicate the
193 prediction of declining V_a for any given trait in small populations, V_a will be reduced on average,
194 especially for traits with primarily additive inheritance; and 2) eventually, the inexorable decline in
195 π in very small populations means that all small populations will eventually lose V_a and their
196 ability to adapt to environmental change. Adaptive potential in such populations will be severely
197 limited unless V_a is replenished by new mutations or migration from differentiated populations
198 (35) (Figure 2).

199 The hypothesis that small populations harbor less V_a has been tested empirically in both
200 laboratory and field settings. Most experimental studies show declines in V_a and weaker responses
201 to selection in small populations or following bottlenecks (53-55). On the other hand, field studies
202 often find a weak association between V_a and genome-wide genetic variation when comparing
203 across populations (48, 56); this weak relationship is likely due to a combination of factors, none
204 of which refute the two takeaways described above.

205 As discussed above, empirical results suggest that V_a may initially increase after a
206 bottleneck due to the conversion of epistatic and dominance variance to V_a (50, 57), and then
207 decline after substantial inbreeding accumulates. Further, V_a is expected to vary among traits and
208 populations depending on genetic architecture, mutation rate, and the mode and history of
209 selection. In practice, most studies are unable to account for these factors and are generally only
210 able to assess a few traits per species/population. Estimates of V_a for each trait are also typically
211 based on a modest number of families. Although the number of traits, populations, and species
212 studied has increased, determining the total V_a for fitness in a given population of conservation
213 concern is not an attainable goal. Additionally, the vast majority of the best-characterized species

214 with respect to V_a in the wild (i.e., most of the species included in (48, 56) meta-analyses) are
215 common. The species and populations in which the relationship between V_a and genetic variation
216 is expected to be strongest, namely, declining species of conservation concern, tend to be most
217 difficult to characterize.

218 Arguably the most important point is that the loss of genetic variation in small and/or
219 bottlenecked populations is inevitable and will eventually lead to reduced V_a and reduce adaptive
220 potential, regardless of short-term and stochastic outcomes. Isolated populations that remain small
221 are unlikely to recover substantial V_a due to the slow rate of mutation and the counteracting loss of
222 variation to genetic drift, and the lack of adaptive potential is problematic for long term viability
223 (43, 44, 47).

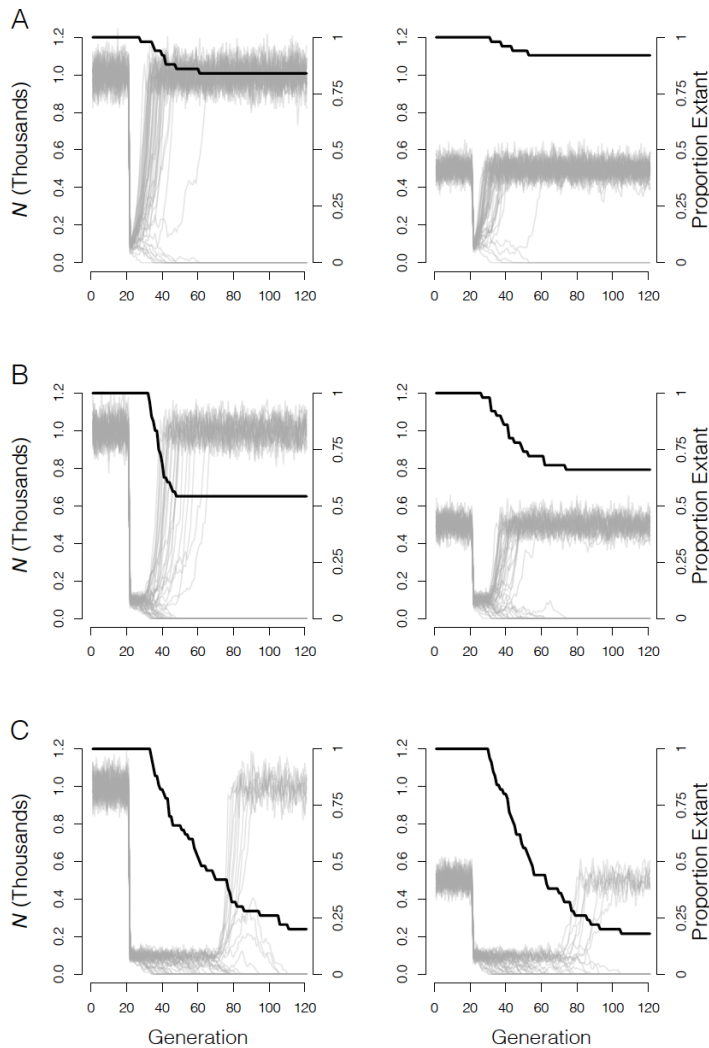
224

225 **3. What is the relationship between genome-wide genetic variation and population viability?**

226 The central question regarding the role of genetic variation in conservation is whether populations
227 with lower π are less likely to persist. Genetic effects on the persistence of a particular population
228 are difficult to predict with certainty because there are many factors involved that are difficult to
229 evaluate, including mating system and demographic history (32, 33), current and future
230 environmental conditions (58), and the extent to which *soft selection* versus *hard selection*
231 predominate (59, 60). Additionally, the highly stochastic demography of small populations, which
232 is exacerbated by inbreeding depression (61), means that widely divergent outcomes can be
233 expected across populations with the same environmental, demographic, and genetic starting
234 conditions. However, theoretical empirical studies have yielded broadly applicable insights into
235 the effects of genetic variation and inbreeding on population viability.

236 Population genetics theory predicts that small, isolated populations with low genetic
237 variation are more likely to go extinct due to genetic effects than larger, more genetically diverse
238 populations under empirically supported mutational assumptions (19, 22, 23, 62). *De novo*
239 mutations following a bottleneck are expected to cause eventual extinction of very small,
240 genetically depauperate populations via *mutational meltdown* (Figure S1) (19). The average time
241 to extinction is shorter under the more realistic scenario where bottlenecked populations carry
242 deleterious mutations at the outset (Figure 3). However, the extinction rate depends strongly on
243 bottleneck duration, with longer restrictions conferring increased extinction due to both
244 demographic stochasticity and the constant increase in drift load. Short-lived bottlenecks are one
245 scenario where viability may sometimes be higher for historically smaller, less genetically diverse

246 populations that have fewer deleterious alleles at the outset of the bottleneck due to historical
247 genetic drift and purging (Figures 1, 3A, 3B). However, this assumes inbreeding depression is the
248 only genetic challenge operating, and simultaneous selection caused by environmental change may
249 reverse this relationship. Longer bottlenecks in isolated populations are expected to result in very
250 high extinction rates due to mutational meltdown regardless of the abundance of deleterious alleles
251 at the outset (19) (Figure 3C).
252



253
254 **Figure 3.** Population viability during bottlenecks from carrying capacity $K=1,000$ (left column)
255 and $K=500$ (right column) to $K=100$. The bottlenecks were 2 (**A**), 10 (**B**), and 50 (**C**) generations
256 in length. The black line shows the proportion of extant populations. Gray lines show population
257 size for each of 50 replicate simulations in each scenario.

258 Empirical studies of population dynamics arguably provide the strongest evidence for the
259 broad benefits of increased genetic variation for population viability. Numerous studies have
260 almost universally found that populations with higher genetic variation have increased population
261 growth and viability (63). For example, lower genetic variation was associated with reduced
262 population growth in alpine ibex (3) and increased local extinction in Glanville fritillary butterflies
263 (2). Inbred laboratory lines of animals, which quickly lose genetic variation, often become extinct
264 substantially more rapidly than control lines (64, 65). Additionally, the infusion of genetic
265 variation via natural (66) and facilitated immigration ('genetic rescue') nearly always increases
266 population growth (35, 36, 67, 68) either by masking of deleterious recessive alleles, or by
267 infusing adaptive genetic variation.

268 The collapse of the Isle Royale wolf population after a mainland male immigrated to the
269 small population has been interpreted as a counter-example to the efficacy of genetic rescue (8).
270 However, detailed documentation indicates that results from this unusual system are unsuitable as
271 a general example of the likely demographic outcome of genetic rescue attempts (67, 69, 70). The
272 immigration of only a single male into Isle Royale makes is unusual in the context of managed
273 genetic rescue attempts which typically involve translocation of multiple individuals into a small
274 population, e.g., (71-73). The single migrant male wolf dominated and increased reproduction,
275 resulting in genetic rescue (an increase in population size following outbreeding). However, his
276 extremely high reproduction resulted in very high inbreeding within two generations and the
277 subsequent dramatic population decline (67, 69, 70). This male was likely just an opportunistic,
278 successful migrant from the nearest population. It is unclear whether he carried an exceptional
279 number of deleterious alleles that drove the subsequent decline, or if inbreeding following
280 exceptionally high reproduction of any individual would have led to a similar demographic
281 outcome.

282 Recovery of some populations from severe bottlenecks, and persistence of some
283 populations despite small N_e and low genetic variation is often cited as a challenge to the idea that
284 low genetic variation and inbreeding reduce population viability (6, 8, 9, 11, 74-77). Soulé (5) [p.
285 178] pointed out the fundamental flaw of this argument, which he referred to as the "fallacy of the
286 accident" nearly 35 years ago: the only observable populations that have experienced bottlenecks
287 are those that survived. The potentially numerous populations that went extinct under similar
288 conditions are unobservable. Counting extant, genetically depauperate populations is therefore an
289 unreliable metric of the extinction risk posed by lost genetic variation and inbreeding. Theoretical

290 population genetics and population ecology both predict that some populations will survive
291 bottlenecks, and some lucky ones will persist for long periods at small population size. However,
292 such cases are likely the rare exception, the lottery winners so-to-speak (5, 67).

293 The most immediate threats to small, genetically depauperate populations are demographic
294 stochasticity and inbreeding depression. However, long term population persistence will in most
295 cases require populations to adapt to environmental change (e.g., climate change, novel diseases,
296 invasive species, etc.) (44, 78). Rapid adaptation to new conditions is possible, but requires
297 sufficient genetic variation and relatively large population size (53, 79). All of the material above
298 highlights the fundamental importance of maintaining large, connected, genetically diverse
299 populations. Long term population viability requires having both manageable *genetic load* and
300 adaptive potential associated with genome-wide genetic variation.

301

302 **4. Simulation-based inferences of the effects of genetic variation and inbreeding on** 303 **population viability**

304 Simulation-based studies showed long ago that inbreeding depression can substantially increase
305 extinction risk (23, 80). However, our increasing understanding of deleterious mutation parameters
306 (e.g., deleterious mutation rates, and the distribution of fitness effects [DFE]) combined with the
307 availability of sophisticated, user-friendly simulation software (81) will likely advance our
308 understanding of inbreeding depression and purging within the field of conservation.

309 While there is much to learn about deleterious mutation parameters, a lot is known about
310 the most important elements. First, deleterious mutations arise frequently (15, 16, 82-84), and
311 large effect deleterious alleles appear to be a major driver of inbreeding depression (14, 85-87).
312 For example, lethal alleles arose via mutation at a rate of $\sim 3\%$ per diploid genome in *Drosophila*
313 (14). Inbreeding depression appeared to be largely due to highly deleterious alleles originating in a
314 subset of pedigree founders in sheep and mice (86, 87). Lethal and other large effect deleterious
315 alleles are frequently observed in small natural populations, humans, and model organisms (14, 83,
316 85, 88-90). The majority of humans and *Drosophila* likely carry one or more recessive lethal
317 alleles (85, 89, 90). Deleterious mutations appeared at a rate of $U=1.2$ /diploid genome/generation
318 in *Drosophila* (15) and $U = 1.6$ in hominids (16). Mutation accumulation studies show that the
319 DFE for deleterious mutations is strongly bimodal, with most mutations having small to moderate
320 effects (e.g. $|s| < 0.25$) and a minority being lethal or semi-lethal (82).

321 Second, the degree of dominance (h) is strongly related to mutation effect size. Direct
 322 observation of dominance effects in yeast and *Drosophila* suggest that nearly neutral deleterious
 323 mutations are slightly recessive on average (h slightly less than 0.5), and highly deleterious
 324 mutations (e.g., $|s| > 0.25$) are nearly fully recessive (h very near zero), with h declining
 325 exponentially as s increases in size (14, 91, 92). There is still much uncertainty regarding
 326 deleterious mutation parameters (see discussion below). However, the best available information
 327 suggests that reasonable values of U are >1 , the DFE is strongly bimodal, and dominance declines
 328 substantially with increasing size of s . These findings guide the simulations presented above
 329 (details in SI).

330 Recently, results from genetically explicit simulations were used to argue that genome-
 331 wide genetic variation is of little importance to population viability, and that purging is likely to
 332 prevent extinction (8, 11, 74). However, these studies excluded large effect deleterious mutations
 333 (Figure S2) and assumed values of U that were between 2.6 and 92.3 times lower than the best
 334 estimate of U in *Drosophila* (Table 1). As a result, these models (8, 11, 74) produce substantially
 335 weaker inbreeding depression (<0.05 to approximately 1 lethal equivalent) than observed in real
 336 populations, where the median number of lethal equivalents for juvenile survival in captive
 337 mammals was 3.1 (93), and 12 for total fitness in wild mammals (23) (Figure S3). There is
 338 substantial uncertainty in deleterious mutation rates, and the DFE, particularly for non-model
 339 organisms. However, the discrepancy between the assumed mutation parameters and the resulting
 340 inbreeding depression in the aforementioned studies (8, 11, 74) and the best available empirical
 341 estimates (Table 1, Figure S3), yield results that underestimate the importance of genetic variation
 342 in conservation, and the efficacy of genetic rescue as a tool in conservation.

343

344 **Table 1.** Deleterious mutation rates used in previous simulation-based analyses of inbreeding
 345 depression and genetic rescue.

Study	Mutation target size	Mutation rate	Proportion deleterious	U^*	$U_{Drosophila}/U^{**}$
Teixeira & Huber (11)	1,000 exons	1×10^{-5} /exon/generation	0.66	0.013	92.3
Robinson et al. (74)	2,000 genes \times 1,000bp	1×10^{-8} /bp/generation	0.7	0.028	42.9
Kyriazis et al. (8)	20,000 genes \times 1,500 bp	1×10^{-8} /bp/generation	0.77	0.462	2.6

346 *U is calculated as $2 \times$ mutation target size \times mutation rate \times proportion of mutations that are deleterious. $^{**}U_{Drosophila}$
 347 is the deleterious mutation rate per diploid genome, $U_{Drosophila} = 1.2$ (15).
 348

349

350

351

351 **5. Is the relationship between genetic variation and conservation status informative of the**
352 **importance of genetic variation to population viability?**

353 It has been suggested that a weak relationship between genetic variation and conservation status
354 (e.g., IUCN Red List) means that genome-wide genetic variation is uninformative of extinction
355 risk (11). However, this relationship is not universally expected, even though extinction risk is
356 strongly affected by genome-wide genetic variation.

357 First, a lag is expected between reduced population size and the loss of genetic variation.
358 Most threatened populations initially decline due to non-genetic factors (e.g., habitat loss, disease,
359 climate change). Thus, multiple generations are required for a substantial reduction in genetic
360 variation, even after severe bottlenecks (Figure 2A). Threatened populations that became small
361 due to non-genetic factors may still have high genetic variation due to this lag. Second, failing to
362 control for other factors that influence genetic variation (e.g., N_e , dispersal, generation time, and
363 mutation rate (11)) can obscure the relationship between genetic variation and conservation status.
364 In contrast, a study controlling for phylogeny (a proxy for the aforementioned confounding
365 factors) showed a significant relationship between genetic variation and conservation status (94).

366 Differences among studies in the measures of genetic variation can further obscure true
367 relationships between genetic variation and conservation status. Estimates of genetic variation for
368 different species used in comparative studies vary widely in the number of sampled individuals
369 and populations, and in the regions of the genome analyzed. Some studies estimate species-wide
370 genetic diversity from a single individual (11, 95, 96) and compare different genetic data types
371 across species (6, 96). Using single genomes to estimate species-wide genetic diversity is
372 problematic because the individuals chosen may not be representative of the species as a whole
373 (e.g., captive individuals (95)). Rather, multiple individuals and populations are necessary to
374 accurately reflect a species' distribution of genetic variation (97, 98). Additionally, estimates of
375 genetic diversity are affected by reference genome quality (99), mapping bias (100, 101), the
376 methods used to measure genetic variation (e.g., whole genome sequencing, RNAseq, RADseq),
377 and bioinformatics approaches (98, 99). Thus, sampling, genetic markers, and analyses should be
378 standardized when measuring the relationship between genetic variation and conservation status.

379 Lastly, IUCN Red List status is an imperfect index of extinction risk because it is a
380 subjective measure of population viability. The IUCN Red List is important for monitoring
381 biodiversity, but the guidelines used to categorize threat levels within the Red List are subject to
382 user interpretation, which can lead to inconsistent assessments (102-106). The imperfect

383 relationship between IUCN Red List status and extinction risk means that Red List status is an
384 inappropriate surrogate for extinction risk in assessing the relationship between genome-wide
385 diversity and extinction risk. Together these issues suggest that the weak relationship between
386 genetic variation and conservation status has little bearing on the importance of genome-wide
387 genetic variation for extinction risk.

388

389 **6. What is the role of functional genetic variation in conservation?**

390 The widespread availability of genomic data for non-model organisms has rapidly advanced our
391 understanding of the genetic basis and evolution of fitness-related traits in natural populations,
392 e.g., (107-111). This revolution has raised the question of how to effectively integrate functional
393 genetic information into conservation practice (112-115). It has repeatedly been suggested that
394 genetic assessment and management of threatened populations should be focused on variation at
395 particular loci that affect particular fitness traits (11, 116-118). However, such gene-targeted
396 conservation approaches are always difficult, and prone to failure for several reasons.

397 First, understanding the genetic basis of fitness remains extremely complicated and
398 challenging (112, 114). While some important traits in natural populations are affected by loci
399 with very large effects, most traits are determined by many small-effect loci (119-121). A
400 comprehensive understanding of the genetic basis of such traits is out of reach for non-model
401 organisms (122). To accurately understand the locus-specific effects on a trait and fitness requires
402 information on dominance and pleiotropy, epistasis, genotype-by-environment interactions, and
403 the amount of linkage disequilibrium with other loci influencing the trait or other fitness
404 components (112). These factors are expected to vary among traits and to differ for the same trait
405 among species and potentially among populations within a species, e.g., (107). Therefore,
406 substantial effort is necessary to understand the conservation relevance of a particular genetic
407 variant and predict whether the benefits of gene-targeted conservation actions outweigh potential
408 detrimental effects (112, 114).

409 A classic example of the potential for undesirable outcomes of gene-targeted conservation
410 management is the suggestion that genetic management of captive and wild populations should be
411 designed around maintaining genetic variation at the major histocompatibility complex (MHC)
412 (11, 116, 117, 123). The MHC has been studied in great detail in humans because of its
413 importance in immunity, organ transplantation, and autoimmune disease, but its organization is
414 poorly understood in most other vertebrates. Although there is strong evidence for its adaptive

415 importance, some variants have detrimental effects, and the adaptive effects of other variants
416 appear to be environmentally dependent (124). Detailed examination of the fitness effects of MHC
417 alleles and haplotypes is necessary to determine how much maintaining MHC variation enhances
418 fitness.

419 Additionally, as highlighted multiple times over the last 35 years (112, 125-129), basing
420 conservation management on a small subset of loci risks increasing the loss of genetic variation
421 elsewhere in the genome. Such efforts would be counterproductive unless the gain in mean fitness
422 associated with gene-targeted management is greater than the loss in fitness associated with lost
423 genome-wide genetic variation (112). This highlights the challenges and pitfalls of gene-targeted
424 conservation. When recommendations for maintaining genome-wide genetic variation versus
425 particular adaptive variants are in conflict, a cost-benefit analysis of the two approaches should be
426 performed and a composite solution identified (112). Recent cases where genomic analyses have
427 revealed that large effect loci play a key role in traits of conservation importance, e.g., (107, 108,
428 110, 130) will be the first to empirically test the efficacy of gene-targeted conservation
429 approaches.

430

431 **Discussion**

432 Genomic data should be used to challenge findings from population genetics theory and previous
433 empirical data that form the basis for genetic management of small populations. Recent genomic
434 studies provide useful fodder to determine how to effectively use genomic data to improve
435 conservation in ways that were previously impossible. Examples are emerging of how
436 understanding functional genetic variation could improve recommendations to conserve imperiled
437 populations (107, 108, 110, 130), making genomic data more useful for conservation than ever
438 before. However, genomic data have not discredited the decades worth of evidence that inbreeding
439 depression, mutational meltdown, and loss of adaptive potential are major threats to conservation.

440 Identifying genetic variants that affect fitness traits undoubtedly advances understanding of
441 the genetic basis of adaptation, and that is important in itself (131). However, placing conservation
442 priority on a small, apparently adaptive portion of the genome ignores what may be the vast
443 majority of variation elsewhere in the genome that will fuel adaptation to unpredictable future
444 conditions (112, 114, 125, 126). This approach is reminiscent of the “adaptationist programme”
445 that Gould & Lewontin (132) criticized >40 years ago for being overly enamored with adaptive
446 explanations for interesting traits (‘spandrels’) without considering that they might have arisen by

447 accident, and that they are but one part of the whole, complex organism (114). Now, as then, we
448 should avoid the temptation to place undue priority on putatively adaptive loci ('molecular
449 spandrels' (133)) without first considering the rest of the genome. Our inability to predict future
450 changes in genotype-by-environment interactions should lead us to recognize the importance of
451 genome-wide genetic variation (including presently neutral variation), and more importantly, the
452 factors that make it possible – large livable habitats and natural patterns of connectivity among
453 them. Conserving genetic variation across the whole genome is almost certainly the most reliable
454 approach to conserve the genetic variation that matters.

455 We know of no convincing evidence that supports abandoning the focus on genome-wide
456 genetic variation in exchange for a focus on functional variation. The recent simulation studies that
457 have been used to discount the importance of genome-wide genetic variation in conservation (8,
458 11, 74) are based on assumptions that are inconsistent with the preponderance of empirical data on
459 the genetics of inbreeding depression and its effect on population viability (see above). Some
460 small populations may not suffer strong inbreeding depression, and some may not rebound
461 following the introduction of genetic variation. However, as pointed out in the formative years of
462 conservation biology, we must resist the temptation to dismiss the extinction risks associated with
463 lost genetic variation in small populations (5).

464 Although population genetics theory has done a remarkably good job of predicting patterns
465 now observable in genomic data, many questions remain unanswered that will improve the utility
466 of genomic data in conservation. For example, how prevalent is soft selection? The presence of
467 soft selection could help explain some of the instances where populations persist for long periods
468 despite inbreeding (59, 60). How much do U and the distribution of fitness effects for deleterious
469 mutations vary among taxa? U may be rather consistent within some taxonomic groups (e.g.,
470 mammals) where the number of genes is strongly conserved (134). Nevertheless, variation among
471 taxa in gene number, mutation rate, and the amount of intergenic DNA that is subject to
472 deleterious mutation is an important consideration for assessing the fitness effects of inbreeding.
473 Lastly, while it is clear that the distribution of mutation fitness effects is bimodal (82),
474 understanding the specific shape of this distribution, and how much this varies among taxa, is
475 important for our understanding of the extinction risks associated with small population size and
476 inbreeding.

477 Genomic data will undoubtedly continue to be used to revisit and refine insights gained
478 since genetics was first applied to conservation and to understand the extinction process (4, 5, 46,

479 135). So far, genomic data have reinforced earlier findings showing that genome-wide genetic
480 variation is key to population viability. Given the increasing rate of habitat loss and fragmentation,
481 failing to recognize and mitigate the effects of lost genome-wide genetic variation would only
482 exacerbate the biodiversity crisis.

483

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491

492 **Data availability**

493 Materials to replicate the simulations are available at <https://doi.org/10.5281/zenodo.5513957>.

494

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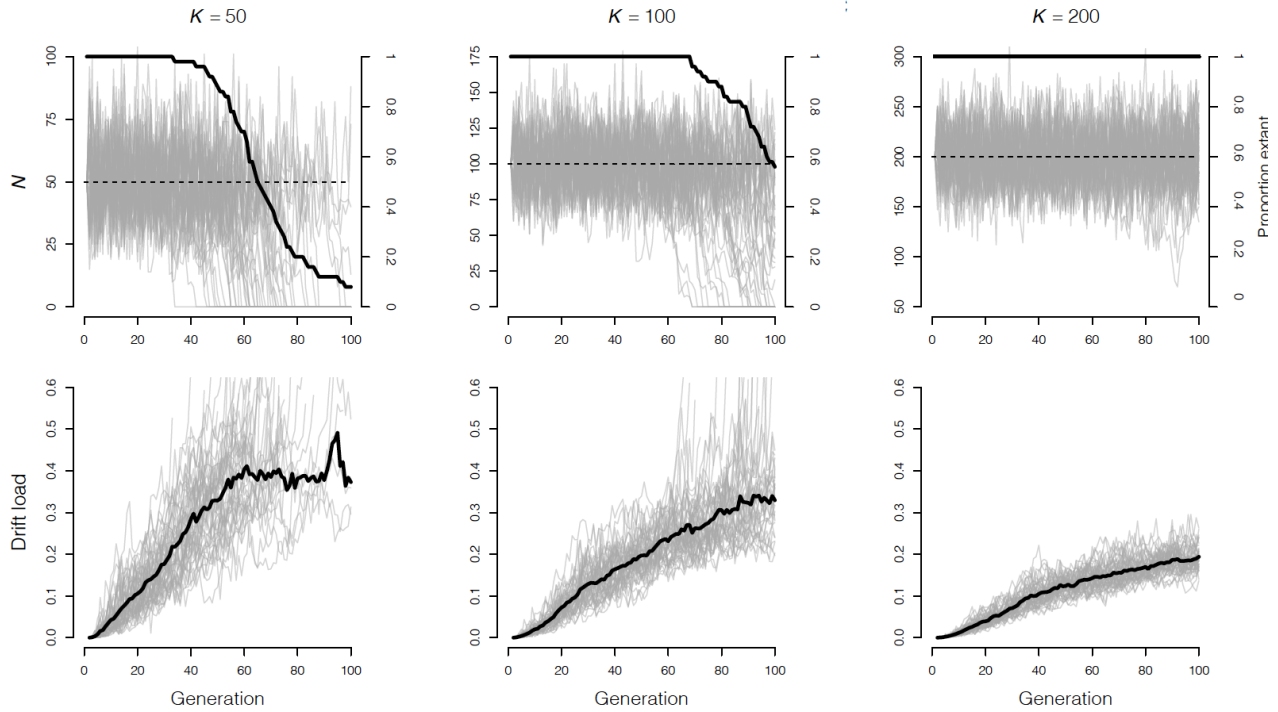
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- 794
795
796

797 **Glossary:**
798 ***Adaptive potential:*** The ability of a population to evolve adaptively in response to selection.
799 Usually measured as narrow sense heritability (the proportion of phenotypic variance attributed to
800 additive genetic effects).
801
802 ***Drift load:*** The reduction in mean fitness of a population due to homozygosity for deleterious
803 alleles.
804 ***F:*** The individual inbreeding coefficient: the identical-by-descent fraction of an individual's
805 genome.
806
807 ***Genetic load:*** The reduction in fitness due to all genetic effects arising from both segregating and
808 fixed deleterious alleles.
809
810 ***Genetic rescue:*** Increase in population growth or reduction in genetic load arising from the
811 immigration of individuals with new alleles.
812
813 ***h:*** The dominance coefficient. A derived allele is recessive when $h=0$ (heterozygous genotypes
814 have the same mean fitness as homozygous wildtypes), and dominant when $h=1$ (heterozygous
815 genotypes have the same mean fitness as homozygous derived allele genotype), and additive when
816 $h=0.5$ (heterozygous genotypes have fitness midway between the alternative homozygous
817 genotypes).
818
819 ***H:*** Heterozygous fraction of an individual's genome.
820
821 ***Hard selection:*** Where an individual's absolute fitness depends only on its phenotype or genotype
822 and is independent of the phenotypes or genotypes of other individuals in the population.
823
824 ***Identical-by-descent:*** Two segments of DNA are identical-by-descent when they both descend
825 from a single haploid genome in a recent ancestor.
826
827 ***Inbreeding:*** Mating between relatives.
828
829 ***Inbreeding depression:*** Reduced fitness of individuals whose parents are related.
830
831 ***Inbreeding load:*** A measure of the potential for inbreeding to reduce fitness, measured by the
832 number of ***Lethal equivalents***, which is a set of alleles that would on average cause death when
833 homozygous.
834
835 ***Mutational meltdown:*** Extinction of a population due to the synergistic interactions of population
836 decline, genetic drift, and the accumulation of deleterious alleles.
837
838 **π :** Nucleotide diversity: expected proportion of nucleotide differences between randomly chosen
839 pairs of haploid genomes in a population.
840
841 ***Purging:*** Increased selective elimination of deleterious, partially recessive alleles that are exposed
842 to purifying selection via inbreeding.

843 ***Soft selection:*** Selection where an individual's fitness depends on its phenotype or genotype
844 relative to others in the same population.

1 SUPPLEMENTARY INFORMATION

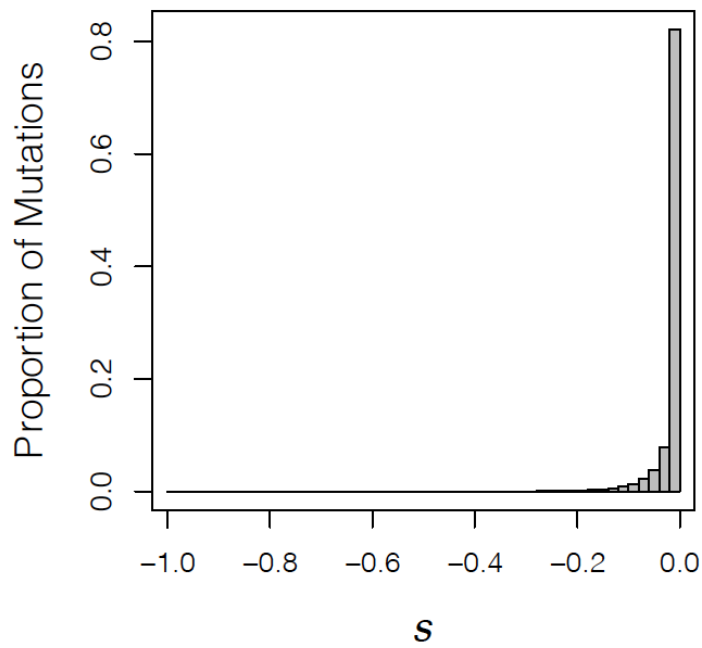
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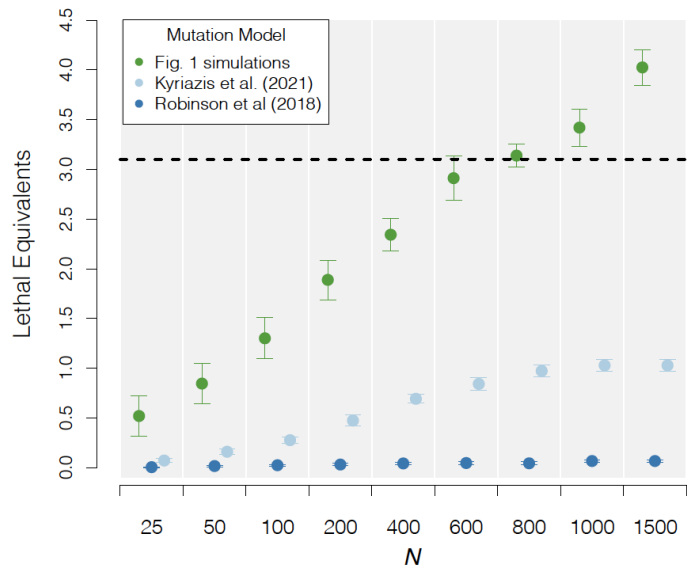
5 **Figure S1.** Mutational meltdown via *de novo* mutation in isolated populations. Panels in the top
6 row show population size N (gray lines, left vertical axis), and the proportion of extant populations
7 (thick black line, right vertical axis) for 50 replicate simulations of populations with carrying
8 capacities (K) of 50 (A), 100 (B), and 200 (C). The bottom row shows the drift load for each
9 simulation replicate (gray lines), and the mean across all non-extinct populations (thick black line).
10 These simulations with hard selection have a ratio of effective population size (N_e) to N of
11 approximately 0.25 on average (Figure S5).

12



13
14 **Figure S2.** Gamma distribution (shape parameter = 0.186 and scale parameter = 0.071) of fitness
15 effects (s) for deleterious mutation assumed in Teixeira and Huber (2021), Robinson et al. 2018,
16 and Kyriazis et al. (2020). Highly deleterious mutations are effectively excluded here (compare to
17 Figure S4 and the results reviewed in Eyre-Walker & Keightley (1)).

18
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24
 25 **Figure S3.** Number of lethal equivalents at approximate mutation-drift-selection equilibrium
 26 under the mutation models of Kyriazis et al. (2), Robinson et al. (3), and the simulation model in
 27 Figure 1 for constant population sizes ranging from $N_e = 25$ to $N_e = 1,500$. The error bars represent
 28 the standard deviation across 10 simulation replicates. The dashed line represents the median
 29 number of lethal equivalents for juvenile survival for captive mammals from Ralls et al. (4). Note
 30 that O’grady et al. (5) estimated an average of 12 lethal equivalents across all fitness components
 31 in wild mammals.

32
 33 **Simulations illustrating the relationship between genetic variation and fitness**
 34 We use individual-based simulations implemented in R (6-9) to illustrate the relationships among
 35 genetic variation, population size, additive genetic variance (V_a), inbreeding load, drift load, and
 36 population viability. These are intended to demonstrate patterns that arise directly from population
 37 genetics theory under empirically supported combinations of the key parameters. The simulated
 38 organism is a self-incompatible hermaphrodite, and has non-overlapping generations, and mean
 39 fecundity of 4 (6) when selection was hard (population size is temporally variable), and 2 when
 40 selection was soft (population size is temporally constant). Details on the implementation of hard
 41 versus soft selection are provided below. Partially recessive deleterious mutations, and mutations
 42 that affect the quantitative trait affect fitness by viability selection before breeding when
 43 population size is temporally variable (selection is hard), and during the reproduction phase when

44 population size is constant (selection is soft). The simulations in Figures 1 & 2 in the main text
45 include both partially recessive mutations (as described below), and mutations that affect a
46 quantitative trait (also described below). The simulations shown in Figures 3 (main text), S1, and
47 S3 include partially recessive deleterious mutations, but do not incorporate selection on a
48 quantitative trait.

49 Simulations with temporally variable population size (Figures 3 & S1) assume a ceiling
50 model of density dependent fitness. Here, when population size is $>$ carrying capacity (K), mean
51 fitness is penalized so that the expected number of offspring forming the next generation is K .

52

53 *Mutations affecting a quantitative trait under stabilizing selection*

54 Our model for the inheritance of a quantitative trait is from Kardos & Luikart (6). The quantitative
55 trait is assumed to have an optimal phenotype value of $\theta = 0$ (in arbitrary units), a per diploid
56 genome per generation mutation rate of $U_q = 0.147$, with phenotypic effects (a) drawn from a
57 uniform distribution ranging from -0.5 to 0.5, an environmental variance of $V_e = 4$. We assume a
58 Gaussian fitness function:

59

$$60 \quad W_{q,i}(z) = W_{\max} e^{-\frac{(z_i - \theta)^2}{2c^2}}, \quad (1)$$

61

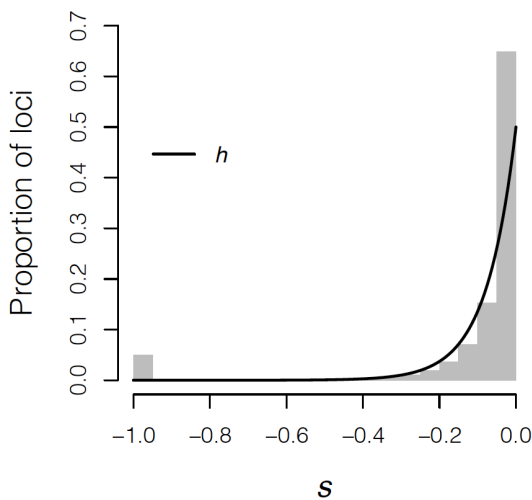
62 where $W_{q,i}(z)$ is the expected fitness of individual i with quantitative trait value z_i , c is the
63 standard deviation of the fitness function [set to $c = 6$ as in (6)], z is the individual's phenotype
64 value, and W_{\max} is the expected fitness of an individual with phenotype of $z = \theta$ and no partially
65 recessive deleterious mutations (set to $W_{\max} = 2.5$). W_{\max} is equivalent to the intrinsic population
66 growth rate for a perfectly adapted population with population size very near zero. Smaller values
67 of W_{\max} (e.g., $W_{\max} = 1.5$) resulted in nearly all large populations going to extinction before
68 reaching mutation-drift-equilibrium for lethal equivalents when selection was hard (see below).

69

70 *Deleterious mutations affecting fitness*

71 Deleterious mutations act directly on individual fitness. We assume a deleterious mutation rate per
72 diploid genome of $U = 1.2$, as observed in *Drosophila* (10), which is substantially lower than in
73 hominids ($U = 1.6$) (11). The location of a mutation is assigned randomly across 38 chromosomes,
74 the number of autosomes in *Canids* (12), each with a 50 cM genetic length. We assume a gamma

75 distribution of mutation fitness effects (s , the expected reduction in fitness for derived allele
76 homozygotes relative to wild type homozygotes), with shape parameter = 0.5 and scale parameter
77 = 0.1, augmented so that 5% of deleterious mutations are lethal (Figure S4). This distribution
78 mimics the distribution of fitness effects observed in mutation accumulation experiments (1), and
79 is consistent with known contribution of both lethal and small-effect, partially recessive mutations
80 in model organisms, humans, and non-model organisms, e.g., (13-15). We assume an exponential
81 model of the relationship between dominance (h) and s as $h = 0.5e^{-13s}$, which closely mimics
82 experimental results in model organisms (16, 17), where mutations with s very near 0 are generally
83 nearly additive ($h \approx 0.5$), and mutations with s near -1 (lethals) are essentially completely
84 recessive ($h \approx 0$, Figure S4). Using the higher deleterious mutation rate of hominids would result
85 in an even larger gap between the resulting fitness effects of inbreeding here compared to Teixeira
86 & Huber (18), Robinson et al. (3), and Kyriazis et al. (2) (Figure S3).



87
88 **Figure S4.** The distribution of selection coefficients (s) for deleterious mutations in our
89 simulations. The black line shows the dominance coefficient h as a function of s .
90
91 The fitness reduction arising from partially recessive deleterious mutations for individual i is
92 calculated as

93
94
$$\Delta W_i = \sum_{j=1}^n \eta_{i,j} \begin{cases} h_j s_j & \text{if } \eta_{i,j}=1 \\ s_j & \text{if } \eta_{i,j}=0 \text{ or } 2 \end{cases}, \quad (2)$$

95

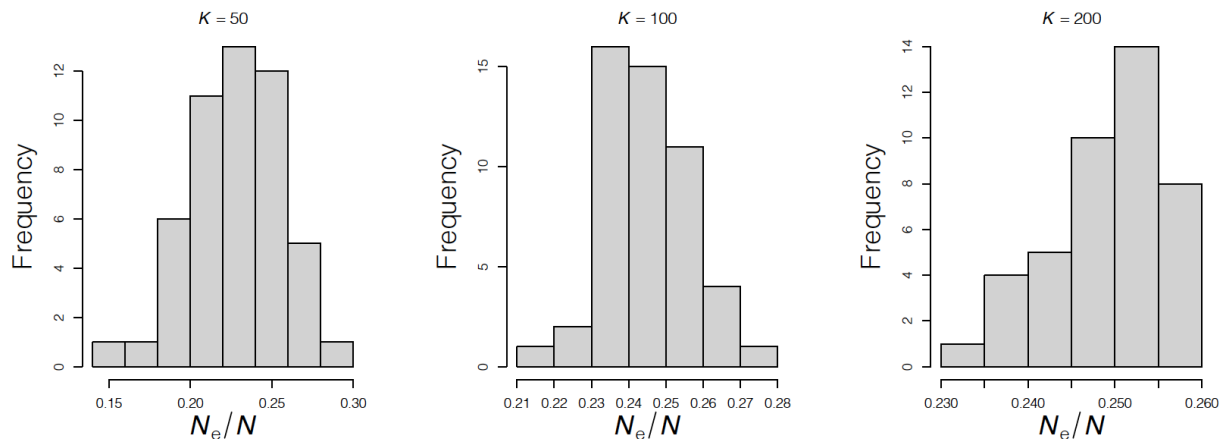
96 where $\eta_{i,j}$ is the count of the derived deleterious allele at the j th of the n loci where there has been
 97 a deleterious mutation in individual i . h_j and s_j are the dominance and selection coefficients,
 98 respectively, at locus j . Subtracting ΔW_i from $W_{q,i}(z)$ (eq. 1) yields the expected fitness of
 99 individual i given the fitness effects of the quantitative trait and partially recessive deleterious
 100 mutations.

101

102 *Hard versus soft selection*

103 Some of our simulations force population size to be constant (Figures 1, 2, S3) to simplify our
 104 analyses of the effects of population size on the parameters of interest. Constant population size
 105 implies that selection on the phenotype and arising from deleterious mutations was soft. Here, the
 106 mean fecundity is by definition 2, such that the population growth rate is exactly $\lambda = 1$, and the
 107 expected fitness of an individual with a particular genotype depends on the genotypes of others in
 108 the population (19). Selection in these cases is implemented during the reproduction phase.

109 Our other simulations allowed population size to fluctuate through time (Figures 3 and S1)
 110 to illustrate genetic effects on population viability. When population size is allowed to fluctuate
 111 through time, selection is hard, where an individual's fitness depends only on its genotype, and
 112 population fitness (population growth rate) depends on the collection of genotypes of all the
 113 individuals in the population (19). Here, selection is imposed via selection on juvenile survival
 114 before the breeding phase.



115
 116 **Figure S5.** Distributions of the ratio of effective population size (N_e) to census population size (N)
 117 in simulations from Figure S1. N_e was calculated as $N_e = (1/\overline{\Delta F})/2$, where $\overline{\Delta F}$ is the mean per
 118 generation change in the pedigree inbreeding coefficient in the population over the first 50
 119 generations of the simulation.

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